



## **An Experimental Study to Evaluate Hepatoprotective Activity of Herbal Formulation in Rats**

**N. A. Khan<sup>1</sup>, M. Nasiruddin<sup>2\*</sup>, I. A. Khan<sup>2</sup>, A. Perveen<sup>1</sup> and A. A. Khan<sup>3</sup>**

<sup>1</sup>Department of Ilmul Advia, A.K. Tibbiya College, Faculty of Unani Medicine, A.M.U, Aligarh, U.P., 202002, India.

<sup>2</sup>Department of Pharmacology, J.N. Medical College, A.M.U, Aligarh, U.P., 202002, India.

<sup>3</sup>Department of Anatomy, J.N. Medical College, A.M.U, Aligarh, U.P., 202002, India.

### **Authors' contributions**

*This work was carried out in collaboration between all authors. Authors NAK, MN and IAK designed the study, wrote the protocol, and wrote the first draft of the manuscript. Author AP managed the literature searches, analyses of the study performed, managed the experimental process and author AAK performed histological examination. All authors read and approved the final manuscript.*

### **Article Information**

DOI: 10.9734/BJPR/2015/18427

#### Editor(s):

(1) Vasudevan Mani, Universiti Teknologi MARA (UiTM), Selangor, Malaysia.

#### Reviewers:

(1) Bhaskar Sharma, Suresh Gyan Vihar University, Jaipur Rajasthan, India.

(2) Mutlu Çayırılı, Mevki Military Hospital, Turkey.

(3) Anonymous, Lagos State University, Lagos, Nigeria.

Complete Peer review History: <http://sciencedomain.org/review-history/10605>

**Original Research Article**

**Received 21<sup>st</sup> April 2015**  
**Accepted 4<sup>th</sup> July 2015**  
**Published 19<sup>th</sup> August 2015**

### **ABSTRACT**

**Objective:** To study the hepatoprotective activity of herbal formulation against CCl<sub>4</sub> induced hepatotoxicity in rats.

**Materials and Methods:** Twenty four Charles Foster albino rats of either sex, weighing 150-200 g were divided into four groups of 6 animals each. The animals of group I and II were administered distilled water in the dose of 1 mL/kg, orally, daily for 8 days. Group III and IV rats were administered Silymarin (100 mg/kg/day) and herbal formulation (150 mg/kg/day) orally, daily for 7 days respectively. On 7<sup>th</sup> day, carbon tetrachloride (CCl<sub>4</sub>) was administered in the dose of 2mL/kg as 1:1 mixture with liquid paraffin i.p. to induce hepatotoxicity in animals of group II, III and IV along with routine treatment. On 8<sup>th</sup> day all animals were sacrificed and blood as well as liver was collected for biochemical parameters and histological examination.

**Results:** There was significant increase in serum glutamic oxaloacetic transaminase (80.2±4.5 vs 26±3.1), serum glutamic pyruvic transaminase (68.7±5.6 vs 24.5±2.6), alkaline phosphatase (53.1±5.6 vs 9.4±2.8) and total bilirubin (2.64±0.09 vs 1.03±0.05) in CCl<sub>4</sub> treated group (P< 0.01)

\*Corresponding author: Email: [naseer\\_bettiah@yahoo.co.in](mailto:naseer_bettiah@yahoo.co.in);

as compared to control group. These parameters were not statistically different in Silymarin and herbal formulation treated groups (SGOT: 47.3±4.2 vs 48.6±3.2, SGPT: 44.2±4.3 vs 46.5±3.5, S.ALP: 23.1±7.4 vs 27.9±2.5 and total bilirubin: 1.68±0.02 vs 1.52±0.05 in Silymarin and herbal formulation group respectively) as compared to control group. But Silymarin and extract treated groups showed significant decrease in these parameters as compared to CCl<sub>4</sub> treated group (P< 0.01). Histology of the liver sections confirmed that the extract prevented hepatic damage induced by CCl<sub>4</sub>.

**Conclusion:** The hydro-alcoholic extract of herbal formulation showed significant hepatoprotective activity against CCl<sub>4</sub> induced hepatotoxicity in rats.

*Keywords: Extract of herbal formulation; Carbon tetrachloride; Silymarin; Hepatoprotective Activity.*

## 1. INTRODUCTION

Liver is one of the most vital organ of the body. The metabolism of various food nutrients, administered drugs and ingested chemical takes place mainly in liver. It converts several toxic substances into nontoxic form and helps in their elimination from the body. The liver has gigantic task of maintaining the body's metabolic homeostasis [1]. This is the most vulnerable organ to be affected by virus, xenobiotics, hepatotoxic drugs (paracetamol, rifampin, isoniazid), excess alcohol ingestion and environmental pollution. Liver diseases are among the most serious ailments. Annually, about 20,000 deaths occur due to liver disease [2]. Hepatotoxic drugs can injure the hepatocyte by generating free-radical or metabolic intermediate that causes peroxidation of membrane lipids and results in liver cell injury. In modern medicine, there is no drug which can prevent or cure hepatic cell injury [3,4].

Numerous medicinal plants and their formulations are used in liver disorders in ethno-medical practice as well as traditional system of medicine in India [5]. In developing countries, more than 80% people depend on traditional system of medicine [6]. Medicinal plants are used in large proportion by Indians because of low cost, easy access and ancestral belief as well as experiences [7]. However a number of herbal formulations have not been still evaluated scientifically for their hepatoprotective effect. Therefore, the study was planned to investigate the hepatoprotective effect of the herbal formulation against CCl<sub>4</sub> induced hepatotoxicity in rats.

## 2. MATERIALS AND METHODS [8]

### 2.1 IAEC Approval

Approval for the study protocol was taken from Institutional Animal Ethics Committee and all

experiments were carried out in accordance to the rules and regulations of IAEC & CPCSEA (Committee for the Purpose of Control and Supervision of Experiments on Animals).

Acute toxicity was carried out and lethal dose was more than 2 g/kg body weight.

### 2.2 Chemicals

The serum glutamic oxaloacetic transaminase (SGOT), serum glutamic pyruvic transaminase (SGPT), alkaline phosphatase (ALP) and total bilirubin (TB) kits were purchased from Siemens, Mumbai. Carbon tetrachloride (CCl<sub>4</sub>) was purchased from Thomas Baker Pvt. Ltd. Mumbai and Silymarin from Sigma-Aldrich, Germany.

### 2.3 Preparation of Extract

All the ingredients (Table 1) of herbal formulation were procured from Dawakhana Tibbiya College, A.M.U., Aligarh and identified by Prof. S. H. Afaq, Department of Ilmul Advia, A.K.T.C., A.M.U., Aligarh, U.P., India. All the ingredients were coarsely powdered and then subjected to extraction. The extraction of powder in hydro-alcohol was done continuously for 6 hours using Soxhlet apparatus. The extract was filtered using Whatman No. 1 filter paper, evaporated on water bath at 40 - 60°C until it dried completely and stored in refrigerator for further use.

The dose of extract for rats was calculated by multiplying its clinical doses described in Unani literature with conversion factor 7 [9].

### 2.4 Animal

Twenty four Charles Foster albino rats of either sex, weighing 150-200 g were procured from Central Animal House of the institute. They were divided into four groups of 6 animals each. They were kept under standard laboratory conditions

and provided commercial diet pellets as well as water *ad libitum*. The room temperature was maintained at 25±1°C.

**Table 1. Ingredients of herbal formulation**

Ingredients	Amount (in gm)
<i>Rheum emodi</i> wall	5
<i>Iris ensata</i> thumb	15
<i>Creteria lacca</i>	5
<i>Cinnamomum cassia</i> blume	5
<i>Rosa damascena</i> mill	10
<i>Crocus sativum</i> linn	3

## 2.5 Treatment

Group I (Control): Distilled water orally in the dose of 1 mL/kg, daily for 7 days.

Group II (Negative control): Distilled water orally in the dose of 1 mL/kg, daily for 7 days.

Group III (Positive control): Silymarin (100 mg/kg) orally daily for 7 days.

Group IV (Test group): Hydro-alcoholic extract of herbal formulation in dose of 150 mg/kg suspended in distilled water orally daily for 7 days.

On 7<sup>th</sup> day, carbon tetrachloride (CCl<sub>4</sub>) was administered in the dose of 2 mL/kg as 1:1 mixture with liquid paraffin i.p. to induce hepatotoxicity in animals of group II, III and IV along with routine treatment. On 8<sup>th</sup> day all animals were sacrificed including group I [10].

## 2.6 Sample Collection

The blood was collected and kept undisturbed for 30 minutes. The blood was centrifuged at 5000 rpm for 15-20 minutes to separate serum. The sera of all animal were estimated for SGOT, SGPT [11], total bilirubin [12] and alkaline phosphatase [13].

## 2.7 Histological Examination

The liver of all rats were removed and fixed in 10% formalin [14]. The tissue was stained with haematoxyline and eosin and histological changes were observed by photomicroscope under high power magnification.

## 2.8 Statistical Analysis

All the results were expressed as mean ± SEM. One-way Analysis of Variance (ANOVA) followed by post-Hoc Dunnett's test was used for the

statistical analysis of data and P < 0.05 was considered significant.

## 3. RESULTS

Administration of CCl<sub>4</sub> to the rats in group II resulted in significant increase in SGOT (80.2±4.5 vs 26±3.1), SGPT (68.7±5.6 vs 24.5±2.6), alkaline phosphatase (53.1±5.6 vs 9.4±2.8) and total bilirubin (2.64±0.09 vs 1.03±0.05) when compared with group I (P<0.01). Administration of silymarin and hydro-alcoholic extract of herbal formulation in group III and group IV respectively prevented the rise in SGOT, SGPT, and TB levels when compared with group II (P<0.01). The levels of liver function test enzymes in the animals treated with the extract were significantly decreased as compared non-treated animals following CCl<sub>4</sub> administration (SGOT: 80.2±4.5 vs 48.6±3.2, SGPT: 68.7±5.6 vs 46.5±3.5, S.ALP: 53.1±5.6 vs 27.9±2.5 and total bilirubin: 2.64±0.09 vs 1.52±0.05 in CCl<sub>4</sub> and herbal formulation group respectively) and were comparable with the standard drug silymarin treated rats (SGOT: 47.3±4.2 vs 48.6±3.2, SGPT: 44.2±4.3 vs 46.5±3.5, S.ALP: 23.1±7.4 vs 27.9±2.5 and total bilirubin: 1.68±0.02 vs 1.52±0.05 in Silymarin and herbal formulation group respectively) (Table 2).

### 3.1 Histological Examination of Liver

Group I: Normal control group showed central blood vessels and radiating cords of hepatocytes as well as the vascular sinusoids with no evidence of fatty changes, necrosis or inflammation (Fig. 1).

Group II: CCl<sub>4</sub> treated animals showed centrilobular (acidophilic) necrosis and vascular congestion (Fig. 2).

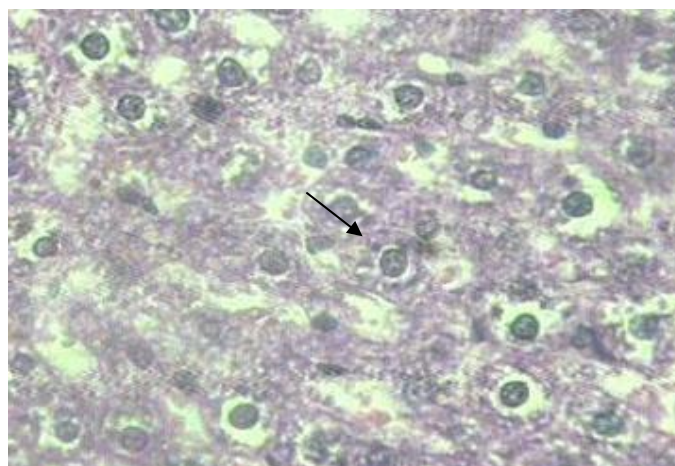
Group III: Silymarin treated rats showed mild vascular congestion and peri-vascular infiltrate of mono nuclear cells and fibroblast. No fatty degeneration was observed (Fig. 3).

Group IV: Hydro-alcoholic extract treated rats showed well preserved liver architecture, only mild vascular congestion and peri-vascular infiltrate of mono nuclear cells and fibroblast and regenerating hepatocytes were observed. The hepatic architecture was found similar to that observed in silymarin treated group (Fig. 4).

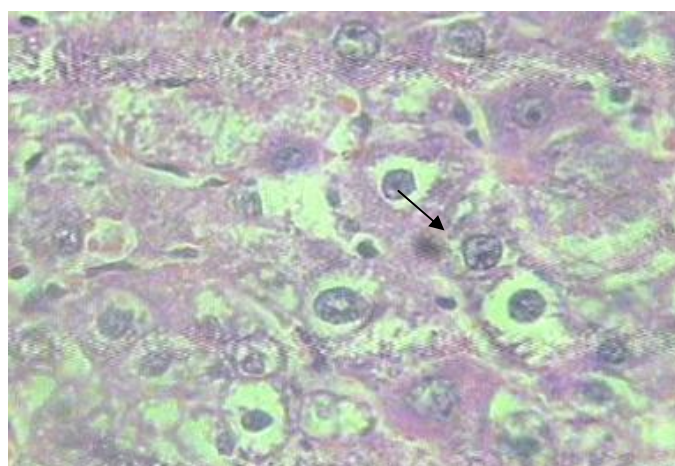
**Table 2. Effect of herbal formulation and silymarin on biochemical parameters of liver function in CCl<sub>4</sub> induced toxicity**

Groups	Treatment	SGPT (IU/L)	SGOT (IU/L)	S.AL.P (IU/L)	Total bilirubin (mg/dL)
GROUP-I Control	Distilled water	24.5±2.6	26±3.1	9.4±2.8	1.03 ±0.05
GROUP-II Negative control	Distilled water + CCl <sub>4</sub>	68.7±5.6*	80.2±4.5*	53.1±5.6*	2.64±0.09*
GROUP-III Positive control	Silymarin 100 mg/kg + CCl <sub>4</sub>	44.2±4.3**	47.3±4.2**	23.1±7.4**	1.68±0.02**
GROUP-IV Test group	Hydro-alcoholic extract 150 mg/kg +CCl <sub>4</sub>	46.5±3.5**	48.6±3.2**	27.9±2.5**	1.52±0.05**

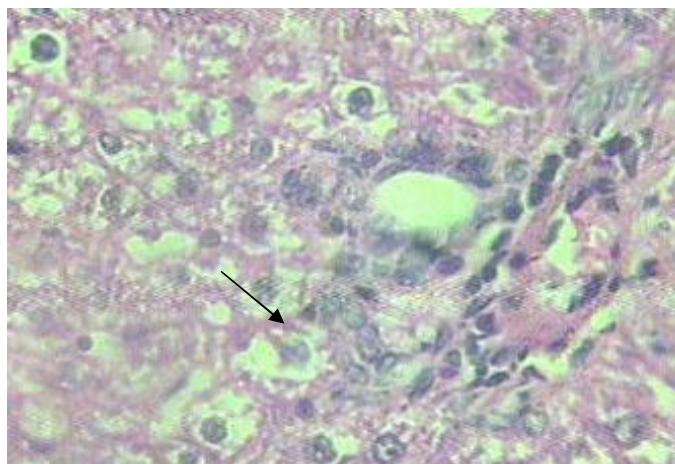
Values are mean ± SEM; N = 6; \*P≤0.01 compared with Group I; \*\*P≤0.05 compared with Group II, SGOT - serum glutamicoxaloacetic transaminase; SGPT- serum glutamic pyruvic transaminase, S.AL.P- Serum alkaline phosphatase



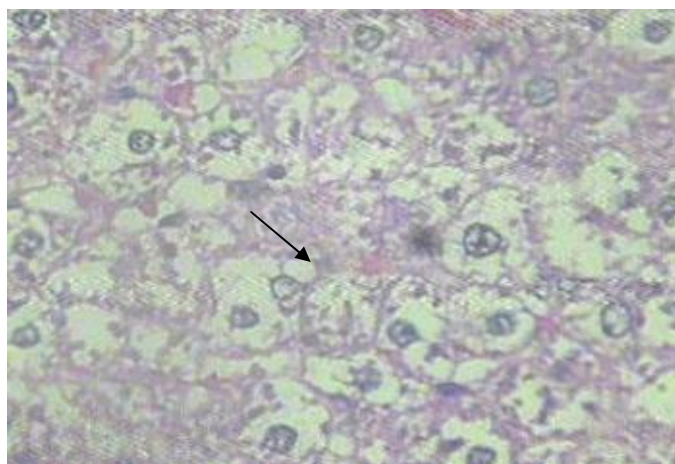
**Fig. 1. Group I- shows central blood vessels and radiating cords of hepatocytes as well as the vascular sinusoids with no evidence of fatty changes, necrosis or inflammation**



**Fig. 2. Group II- shows centri-lobular (acidophilic) necrosis and vascular congestion**



**Fig. 3. Group III-shows mild vascular congestion and peri-vascular infiltrate of mono nuclear cells and fibroblast. No fatty degeneration was observed**



**Fig. 4. Group IV-shows well preserved liver architecture, only mild vascular congestion and peri-vascular infiltrate of mono nuclear cells and fibroblast and regenerating hepatocytes were observed**

#### 4. DISCUSSION

The damage produced by  $\text{CCl}_4$  is described to be similar to the pathological changes seen in infective hepatitis and in many other liver diseases [15]. The liver function test was used to assess the extent of liver damage and the protection induced by the test drug. Silymarin (100 mg / kg) was used as standard hepatoprotective agent for confirming integrity of test system and to compare the efficacy of the test drug as it has been used in the treatment of hepatic diseases [16]. The hepatoprotective properties of Silymarin have been related to inhibition of lipid peroxides formation or

scavenging of free radicals [17]. Carbon tetrachloride ( $\text{CCl}_4$ ) produces hepatotoxicity in a wide variety of mammals. It has been emphasized that  $\text{CCl}_4$  by itself is not biologically active but gets converted to its metabolite – trichloromethyl radical ( $\text{CCl}_3^*$ ) by microsomal enzymes namely CYP2E1, CYP2B1 or CYP2B2 and CYP3A and this is toxic to hepatocytes. This radical react with oxygen to form trichloromethylperoxy radical  $\text{CCl}_3\text{OO}^*$ , which initiates the chain reaction of lipid peroxidation. This brings changes in permeability of mitochondria, endoplasmic reticulum and plasma membranes resulting in elevated levels of transaminases, alkaline phosphatase and

bilirubin. CCl<sub>4</sub> is commonly used for inducing liver toxicity experimentally. This toxic chemical causes peroxidative degradation in liver tissue resulting in coagulative necrotic changes in hepatocytes. CCl<sub>4</sub> produces change around central vein in the liver (Fig. 2) and other oxidative damages resulting in leakage of marker enzymes like SGOT, SGPT and ALP in serum and increase in total bilirubin levels [18,19]. The herbal extract lowered various biochemical markers which shows that they prevent the oxidative damage. Administration of extracts showed significant hepatoprotective activity, which was comparable with silymarin. The qualitative phytochemical studies of hydro-alcoholic extract of herbal formulation also showed positive for flavonoids. Previous studies suggests that plant flavonoids possess antioxidant properties [20] and hence proved useful in the treatment of hepatic damage. The results indicate that the hydro-alcoholic extract of herbal formulation has significant hepatoprotective activity which may be due to higher content of flavonoids like rutin, luteolin and apigenin. Other investigators have screened the hepatoprotective activity of rutin, which is also claimed to have free radical scavenging property and it inhibits lipid peroxidation against CCl<sub>4</sub>-induced hepatic toxicity [21].

## 5. CONCLUSION

The hydro-alcoholic extract of herbal formulation showed significant hepatoprotective activity against CCl<sub>4</sub> induced hepatotoxicity in rats.

## CONSENT

It is not applicable.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Kumar V, Cortan RS, Robbin SL. Robbin's Basic Pathology, 7<sup>th</sup> Ed, 2003; Pennsylvania Saunders; 11, 600.
2. Sharma SK, Ali M, Ansari SH, Gupta J. Evaluation of Indian herbal Hepatoprotective drugs. Hamdard Medicus. 2000;XLIII(2):39-58.
3. Meyer SA, Kulkarni AP. Hepatotoxicity, In: Hodgson E, and Smart RC, Introduction to biochemical toxicology, 3<sup>rd</sup> edition, John Wiley and Sons, New York. 2001;487-490.
4. Harsh Mohan. Text book of Pathology, 4<sup>th</sup> Ed, Jaypee Publication, New Delhi. 2002;569-630.
5. Subramoniam A, Pushpangadan P. Development of phytomedicines for liver disease. Indian Journal of Pharmacology. 1999;31:166-175.
6. Sivalokanathan S, Ilayaraja M, Balasubramanian MP. Efficacy of *Terminalia arjuna* (Roxb.) on N-nitrosodiethylamine induced hepatocellular carcinoma in rats. Indian Journal of Experimental Biology. 2005;43(3):264-267.
7. Ahmad I, Mehmood Z, Mohammad F. Screening of some Indian medicinal plants for their antimicrobial properties. Journal of Ethnopharmacology. 1998;62(2):183-193.
8. Unkeshwar P, Nasiruddin M, Fayazuddin M, Khan Ra, Khan Aa, Tajuddin. Evaluation of Hepatoprotective Activity of *Berberis aristata* against carbon tetrachloride induced hepatotoxicity in rats. Int J Pharm Pharm Sci. 2013;5(4):107-110.
9. Dhawan BN. Organization of Biological Screening of Medicinal Plants with special reference to C.D.R.I programmers. Appendix-I, Lectures UNESCO-CDRI workshop on the use of Pharmacological Techniques for Evaluation of Natural Products, CDRI, Lucknow. 1982;61.
10. Idris Turel, et al. Hepatoprotective and anti-inflammatory activities of *Plantago major* L Indian J Pharmacol. 2009;41(3): 120-124.
11. Reitman S, Frankel S. A colorimetric method for the determination of Serum Glutamic Oxaloacetic and Glutamic Pyruvic Transaminases. Am J Clin Path. 1957;28:56-63.
12. Malloy HT, Evelyn KA. Estimation of Serum Bilirubin Level. J Biol Chem. 1937;119:481-484.
13. Walton H. Marsh, Benjamin Fingurhut & Elaine Kirsch, Alkaline phosphatase estimation; Clinical Chemistry. 1959;5:119-126.
14. Mukherjee KL. Medical Laboratory Technology. Tata McGraw Hill, Publishing Company. 1988;3:1111-1124.
15. Berger BML, Comber H, Esta BB. CCl<sub>4</sub> induced toxicity in isolated hepatocytes, The importance of direct solvent injury. Hepatology. 1986;6:325-327.

16. Choski S, Patel SS, Saluja AK. Silymarin: A promising herbal hepatoprotective drug. *Indian Drugs*. 2000;37(12):566-569.
17. Ferenci P, Dragasics B, Dittrich H. Randomized controlled trial of Silymarin treatment in patients with cirrhosis of the liver. *Journal of Hepatology*. 1989;9:105-113.
18. Reznagel RO. Carbon tetrachloride hepatotoxicity status and future prospects. *Pharmacol Sci*. 1983;4:129-31.
19. Okuno H, Hazama H, Muraze T, Shiozaki Y, Sameshima Y. Drug metabolizing activity in rats with chronic liver injury induced by carbon tetrachloride: Relationship with the content of hydroxyproline in the liver. *Japanese J Pharmacology*. 1986;41:363-71.
20. Hesham RE, Shgeru N. Chemistry of Bioflavonoids. *Indian J Pharm Educ*. 2002;36:191-4.
21. Khalid HJ, Sheikh AS, Anwar HG. Protective effect of rutin on paracetamol and CCl<sub>4</sub> induced hepatotoxicity in rodents. *Fitoterapia*. 2002;73:557-63.

© 2015 Khan et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Peer-review history:*  
*The peer review history for this paper can be accessed here:*  
<http://sciencedomain.org/review-history/10605>